

Non-Class A templates in homology modeling of Metabotropic Glutamate Receptor 2

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Introduction

Homology modeling of Class C GPCRs is a challenging task. Until recently, the only available crystal templates were those of Class A receptors, rather evolutionary distant ones (with sequence identity/similarity oscillating around 19/30 percent) with both ortho- and allosteric binding sites located within heptahelical transmembrane bundle and extracellular loops. However, during the last few months three new crystals appeared, being first structures of GPCRs from other classes of the superfamily. Smoothed Receptor [Wang, 2013], Glucagon Receptor [Siu, 2013], and Corticotrophin-Releasing Factor Receptor 1 [Hollenstein, 2013] potentially open a new avenue for homology modeling studies over distant targets, including Class C GPCRs. In this research for each of those crystal templates a series of 100 homology models of mGlu2 Receptor is generated and evaluated in Virtual Screening like protocol to estimate their applicability for the screening purposes.

The mGluR family consists of eight proteins divided into three groups corresponding to sequence similarities, pharmacology and physiological role. These groups are: I (mGluR1, -5), II (mGluR2, -3) and III (mGluR4, -6, -7, -8). Group II lies in field of our interest due to its potential as therapeutic target for stroke and pain drugs.

Methods

The sequence alignment of the templates and mGluR2 were developed using structural alignment between each crystal and the structure of bovine rhodopsin (PDB: 1u19), and existing alignment of mGluR2 and the latter. Manual adjustments were applied to ensure proper orientation of the amino acids involved in ligand binding [Gregory, 2010].

Homology models were generated with Modeller 9.12 [Sali, 1993] with restrains for the secondary structure of the helices (excluding bending regions).

Docking experiments were conducted with Glide 5.7 (SP mode) using a set of 112 known allosteric modulators of mGluR2 acquired from ChEMBL database and a set of 1000 drug-like decoys from Schrödinger. The analysis of the docking results was performed with home made scripts and Schrödinger Suite 2013.

Results

For all of the investigated templates, a sequence alignments with mGluR2 were created (Fig. 1). Unfortunately for the Smoothed Receptor, the resulting models were not in accordance with the mutational data for mGluR2 and so those models were rejected from further studies. Docking experiments performed led to up to 110 out of 112 active compounds docked for the Glucagon Receptor template and 100 for CRF Receptor 1 mold. Shapes of received binding sites are presented on Figure 2, and enrichment curves obtained for docking active compounds along with decoys are on Figure 3.

Conclusions

The results of VS like experiments show the potential of models on non Class A templates for further research. Satisfactory compatibility with mutational data, along with high number of known ligands recognized prove, that new Class B GPCR crystal templates are indeed of better use for distant targets such as mGluR2.

CRFR1_HUMAN	115	HYHVAAIINYLGHICISLVALLVAFVFLRAR	145	CRFR1_HUMAN	266	VYTDYIQGPMALVLLINFLFLNIV	291
GLR_HUMAN	136	MYSSFQVMYTVGYSLGALLLAILGGLS	164	GLR_HUMAN	300	NMGFWWILRFPVFLAILINFFIVRI	325
OPSD_BOVIN	34	PWQFSMLAAYMFLIMLGFIPINELTYVTQ	64	OPSD_BOVIN	200	NESFVIYMFVHFIIPLIVIFPCYQ	225
GRM2_HUMAN	561	IRWGDARVAVGPVTIACLALATLFLVGVFVR	591	GRM2_HUMAN	722	NHRDASMLGSLAYNVLLIALCTLYAF	747
CRFR1_HUMAN	150	LRNIIHANLIAAFILRNATWVFWQLTMSPE	179	CRFR1_HUMAN	303	TSETIQARKAVKATLVLLPLLGITYMLAFVN	333
GLR_HUMAN	171	CTRNAIHANLFAFVFKASSVLDGLLRT	200	GLR_HUMAN	342	DYKFRILAKSTLTLLIPLLVGHEVFAFVDEHA	373
OPSD_BOVIN	71	PLNYILLNLAVADLFMVFGGFTTLYTSLH	100	OPSD_BOVIN	246	AEKEVTRMVIIMVIAFLICWLPYAGVAFYIFT	277
GRM2_HUMAN	597	VVKASGRELCYILLGGVFLCYMTFFIFIAK	626	GRM2_HUMAN	754	ENFNEAKFIGFTMYTTCIIWLAFLPIFYVTS	785
CRFR1_HUMAN	184	NVGCWRLVTAAYNYFHVNTFFWFMFGEGCYLHTAI	217	CRFR1_HUMAN	338	EVSRRVFIYFNAPLESFQGFVSVFAC	364
GLR_HUMAN	216	LSDGAVAGCRVAAVFMQYGIIVANYCWLVEGLYL	249	GLR_HUMAN	375	GTLRSKAKLFFDLFLSSFGLLVAVLYC	401
OPSD_BOVIN	106	GPTGCNLEGGFATLGGETALWLSLVLAIERVYVV	139	OPSD_BOVIN	285	PIFMTIPAFFAKTSAVYNPVIYIMNK	311
GRM2_HUMAN	629	TAVCTLRRLGLGTAFCVCSYALLTKTNRIARIFG	662	GRM2_HUMAN	794	MCVSVSLSGSVVLGCLFAPKLIILFQ	820
CRFR1_HUMAN	227	RLRAWMFICIGWVPPPIIVAWAI	248				
GLR_HUMAN	263	FFSLYLGIWGAAPMLFVVPWAVK	286				
OPSD_BOVIN	150	ENHAIMGVAFTWVMALACAAPLV	173				
GRM2_HUMAN	677	ASQVAICLALISGQLLIVVAVLVV	700				

Figure 1. Sequence alignment between mGluR2 and Bovine Rhodopsin, Corticotrophin-Releasing Factor Receptor 1 and Glucagon Receptor with indicated point mutation for mGluR (red) and CaSR (blue). Residues in bold correspond to X.50 in Ballesteros – Weinstein notation. Helices I-IV – left panel, V-VII – right panel.

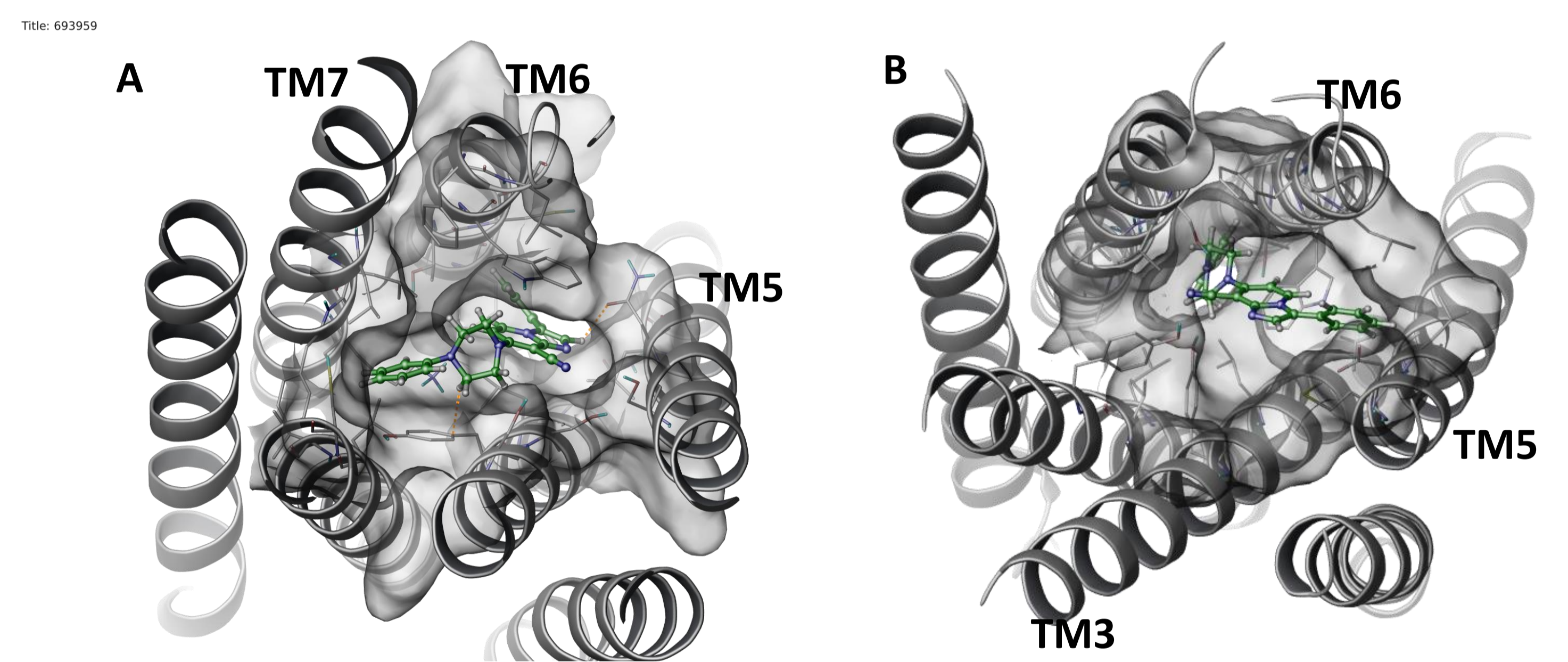


Figure 2. Binding sites of models obtained for different templates: CRFR1 (A) and GLR (B).

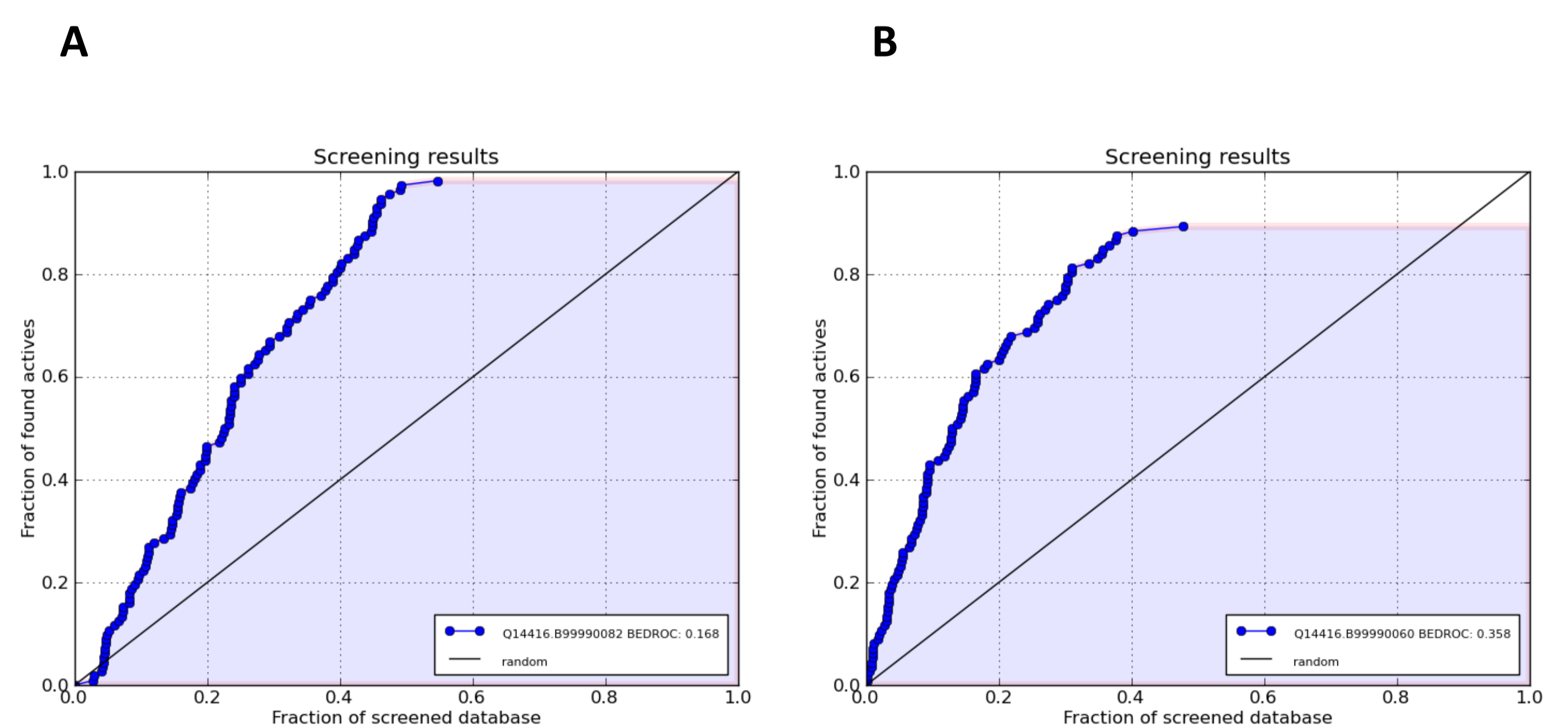


Figure 3. The results of screening like experiments for mGluR2 models built with different templates: CRFR 1 (A) and Glucagon Receptor (B).

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