

# Pharmacological characterization of zinc interaction with 5-HT<sub>7</sub>

Grzegorz Satała, Beata Duszyńska, Andrzej J. Bojarski

Department of Medicinal Chemistry, Institute of Pharmacology Polish Academy of Sciences,

12 Smętna Street, 31-343 Kraków, Poland

e-mail: grzegorz.satala@gmail.com

## INTRODUCTION

Zinc, as an essential trace element in living organisms, has many functions, including participation in various processes within the central nervous system [1]. The role of zinc in depression and its therapy is emphasized by numerous preclinical and clinical studies, however, the exact mechanism of its action is still not fully understood [3]. Our interests are focused on its effects mediated by serotonin receptors, which are key players in the etiology of anxiety and mood disorders [2].

The main objective of this study was to investigate the effect of zinc on the serotonin receptor 5-HT<sub>7</sub> using *in vitro* methods [4,5]. At first, saturation binding assays were performed in a presence of various zinc concentrations in order to determine whether the shift in radioligand affinity reflects an allosteric mode of action. Two different radioligands (of agonistic and antagonistic activity) have been used, as allosteric regulation is highly sensitive to the type of orthosteric ligand (probe-dependence). Next, kinetic effects on radioligand dissociation rate ( $K_{off}$ ) were measured to quantify allosteric effects of zinc ions.

## MATERIALS and METHODS

**Chemicals** [<sup>3</sup>H]5-CT (spec. act. 39.2 Ci/mmol), [<sup>3</sup>H]SB-269970 (62.7 Ci/mol), were purchased from PerkinElmer. Other chemicals were obtained from commercial sources and were of analytical grade.

**Biological material** Membranes were prepared from human embryonic kidney (HEK) 293 cells stably transfected with human 5-HT<sub>7</sub> receptor cDNA.

**Ligand binding assays** For [<sup>3</sup>H]5-CT and [<sup>3</sup>H]SB-269970 bindings crude membrane preparations were incubated in two volumes of assay buffer (0.25 ml) in 37°C for 1 h. Composition of assay buffer was: 50 mM Tris-HCl pH 7.7, 5 mM MgCl<sub>2</sub>, 0.1 mM pargyline and 0.5 mM ascorbic acid. Non-specific binding was defined with the use of 10 μM serotonin. The incubations were terminated by the rapid filtration through Unifilter GF/B plates (PerkinElmer) and subsequent washing with ice-cold buffer using Unifilter harvester. Scintillation cocktail was added and the radioactivity determined in scintillation MicroBeta counter.

**Analysis of data** Analysis of the saturation binding data obtained for the agonist [<sup>3</sup>H]5-CT and antagonist [<sup>3</sup>H]SB-269970 using the program GraphPad PRISM, according to equations [6-9]:

1. „Affinity-shift”-1=[A](1-α)/(α[A]+K<sub>A</sub>)
2.  $pK_{DApp} = -\log([A] + 10^{\log K_A}) + \log(\alpha[A] + 10^{\log K_A}) - \log d$

where  $\log d$  is a fitting constant;  $K_A$  denotes the affinity of zinc for the allosteric site;  $[A]$  the Zn<sup>2+</sup> concentration;  $\alpha$  defines the cooperativity factor, the magnitude by which the equilibrium dissociation constant of either ligand for its site on the receptor is modified by the concomitant presence of the other ligand. Values of  $\alpha$  less than 1 (but greater than zero) denote negative cooperativity, values greater than 1 denote positive cooperativity, and values not significantly different from 1 indicate neutral cooperativity.

## RESULTS

Analysis of saturation isotherms obtained for seven increasing concentrations of zinc (10 μM — 5 mM) [Fig 1. and Fig 2.] revealed decrease in radioligand binding (increased  $K_d$  values in relation to  $K_d$  of [<sup>3</sup>H]5-CT and [<sup>3</sup>H]SB-269970 binding without zinc) which achieved plateau at the highest concentration of zinc used (5 mM). The mechanism by which zinc inhibits the binding of radioligands was further evaluated by a Schild-type plot analysis. Data showed a negative cooperativity of zinc, which indicates allostery.

In order to further evaluate the nature of the interaction of Zn<sup>2+</sup> with the 5-HT<sub>7</sub> receptor, dissociation kinetic assays for [<sup>3</sup>H]5-CT (0,5 nM) in the absence and presence of 500 μM Zn<sup>2+</sup> were carried out [Fig 3.]. The dissociation rate ( $K_{off}$ ) kinetics for [<sup>3</sup>H]5-CT binding to the 5-HT<sub>7</sub> receptor was assayed at 37°C by addition of serotonin (10 μM) to prevent the reassociation. The presence of 500 μM Zn<sup>2+</sup> produced a significant increase of the dissociation kinetic rate of [<sup>3</sup>H]5-CT. The effect of zinc upon the dissociation kinetic rate indicates that the noncompetitive interaction of Zn<sup>2+</sup> at 5-HT<sub>7</sub> receptor may be classified as negative allosteric modulation.

Figure 4 shows results of control experiment, i.e. the screening of other ions on [<sup>3</sup>H]5-CT binding to 5-HT<sub>7</sub> receptor. The specific action of zinc, compared to other ions, is evident.

Summing up, the *in vitro* experiments clearly showed that Zn<sup>2+</sup> ions act as negative allosteric modulator (NAM) at 5-HT<sub>7</sub> receptors.

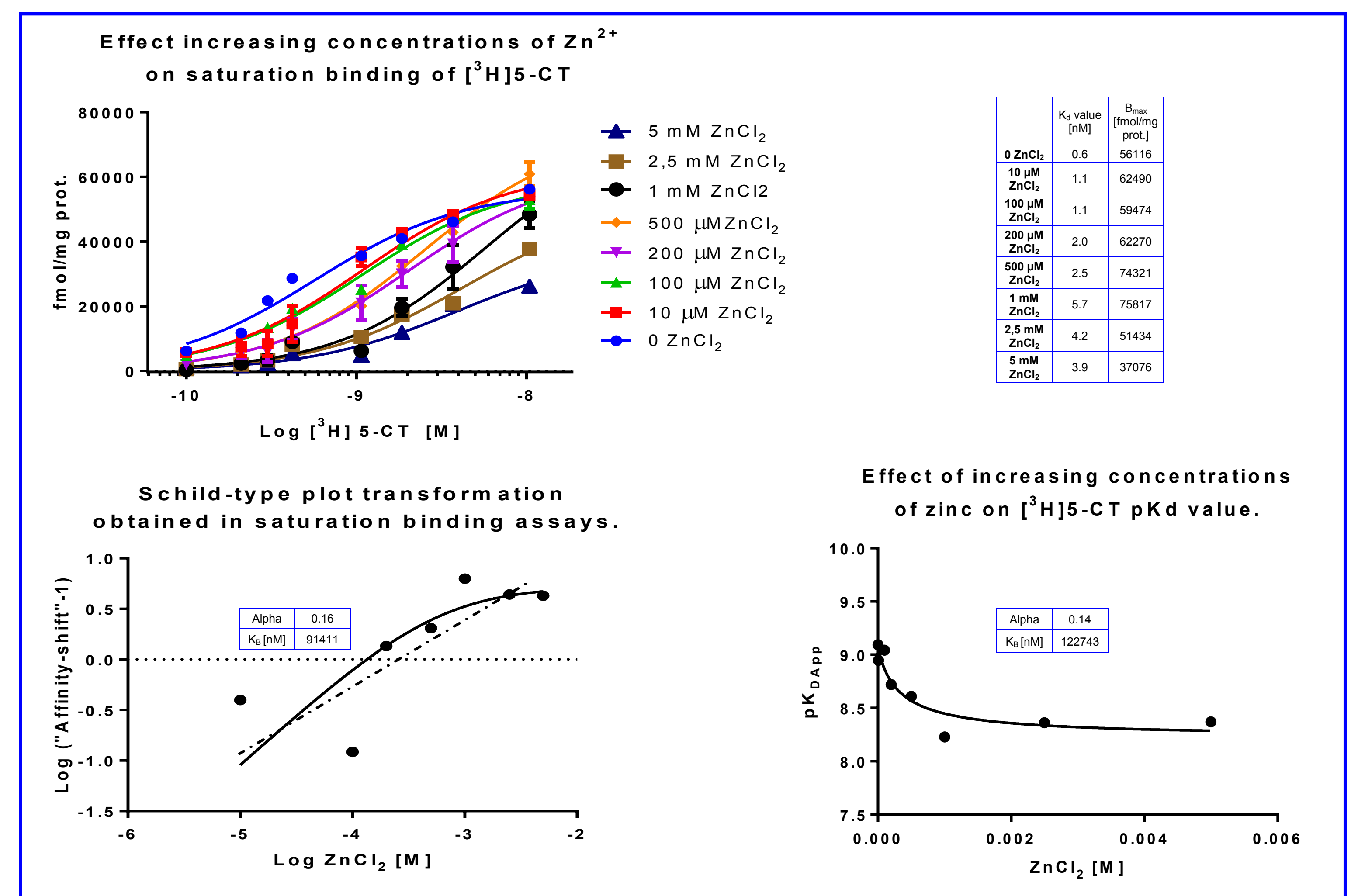


Figure 1.

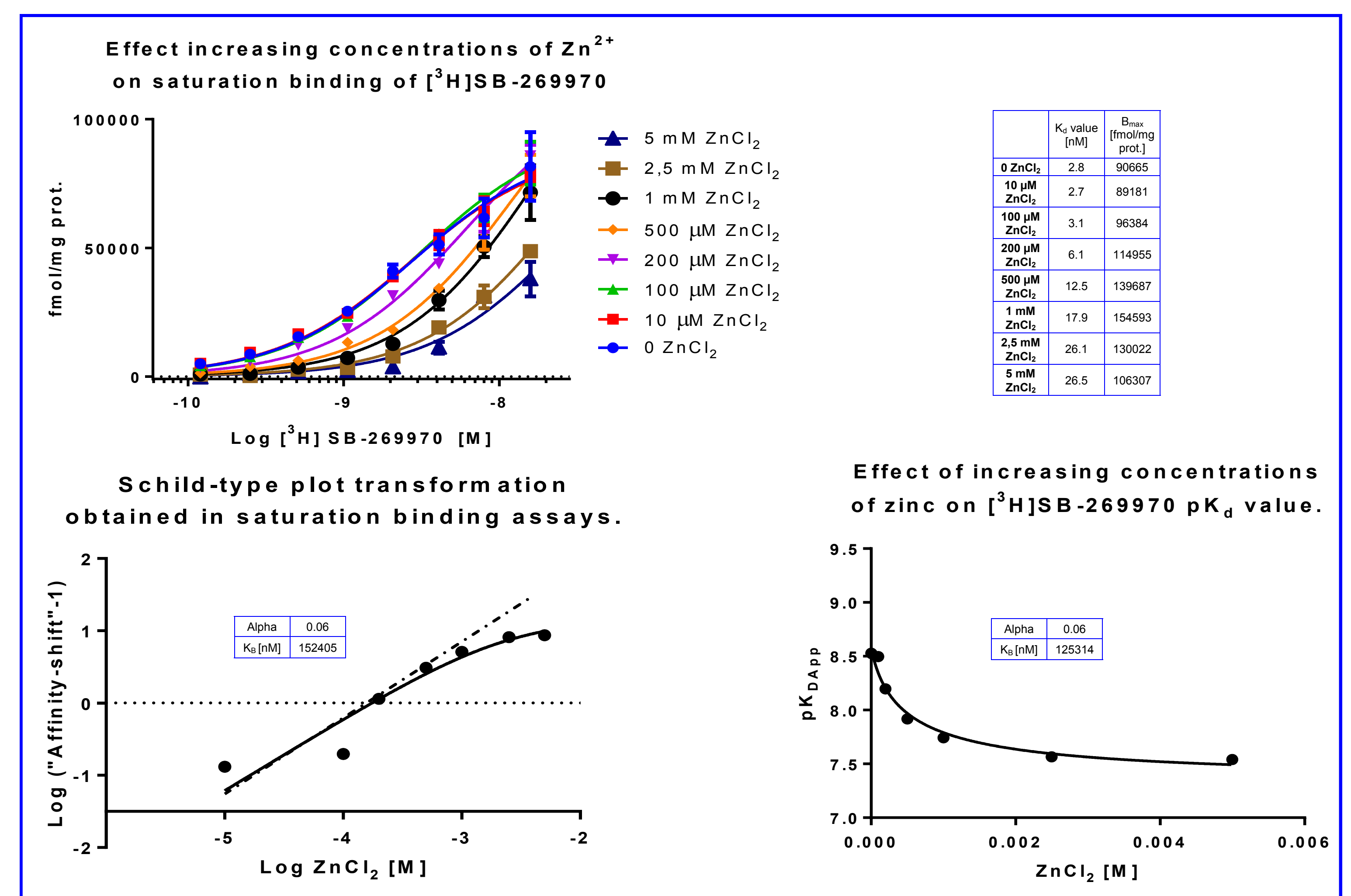


Figure 2.

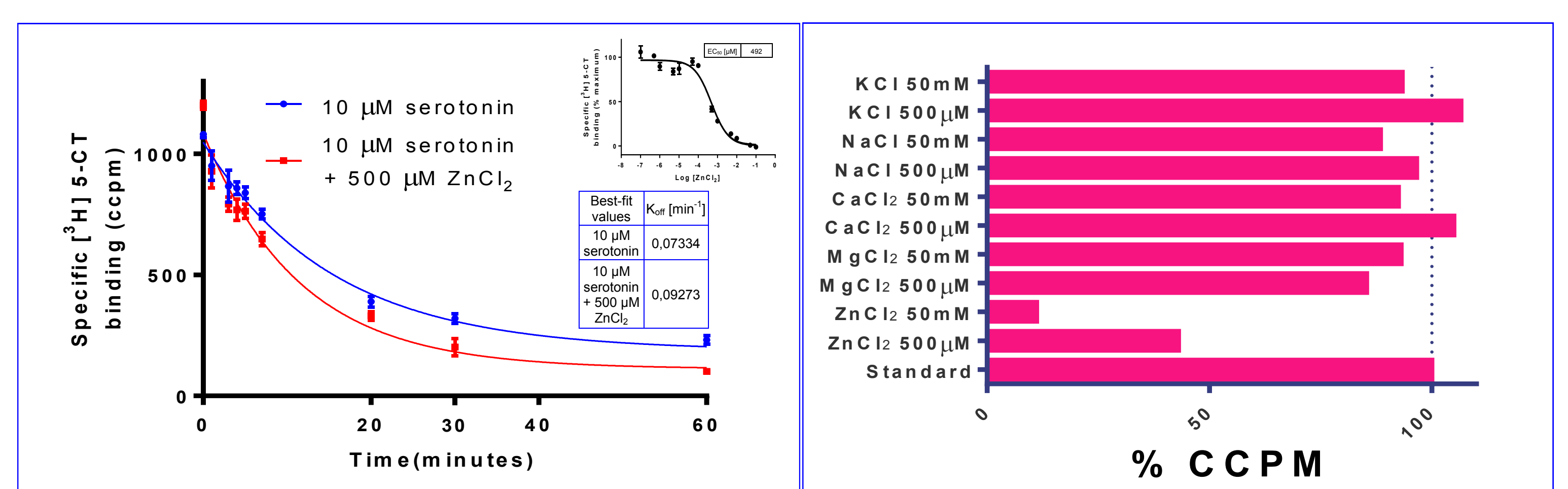


Figure 3.

Figure 4.

## References:

- [1] Chasapis CT, Loutsidou AC, Spiliopoulou CA, Stefanidou ME. Zinc and human health: an update. Arch Toxicol. 2012, 86, 521-34.
- [2] Nichols DE, Nichols CD. Serotonin receptors. Chem Rev. 2008, 108, 1614-41.
- [3] Dickerman B, Liu J. Do the Micronutrients Zinc and Magnesium Play a Role in Adult Depression? Topics in Clinical Nutrition. 2011, 26, 257-267.
- [4] Leach K, Christopoulos A. Quantification of allosteric interactions at G protein-coupled receptors using radioligand binding assays. Curr Protoc Pharmacol. 2011, Chapter 1, Unit 1.22.
- [5] Lazareno S. Determination of allosteric interactions using radioligand-binding techniques. Methods Mol Biol. 2004, 259, 29-46.
- [6] Christopoulos A1, Mitchelson F. Application of an allosteric ternary complex model to the technique of pharmacological resultant analysis. J Pharm Pharmacol. 1997 Aug;49(8):781
- [7] Ehlerl, F.J., 1988. Estimation of the affinities of allosteric ligands using radioligand binding and pharmacological null methods. Mol. Pharmacol. 33, 187-194.
- [8] Christopoulos, A., Kenakin, T., 2002. G protein-coupled receptor allostery and complexing. Pharmacol. Rev. 54, 323-374.
- [9] May, L.T., Leach, K., Sexton, P.M., Christopoulos, A., 2007. Allosteric modulation of G protein-coupled receptors. Annu. Rev. Pharmacol. Toxicol. 47, 1-51

## Acknowledgements:

The study is supported by a grant PRELUDIUM  
DEC-2012/05/N/NZ7/02110 financed by the National Science Centre

